



"WEAK" AND "WEIRD" BANDS OF IMMUNOFIXATION
Part III

In the first two parts of this series of articles on marginal and atypical immunofixation patterns you learned about weak bands and restrictions representing the low concentration extremes of monoclonal components (Part I) and the real and apparent free light chains (Part II). The concluding part of this series will deal with complications certain physico-chemical properties of immunoglobulins can cause. And, of course, there will be a few words about atypical cases that do not fit into any other category.

Polymers, complexes, aggregates and high Ig concentrations

False bands might appear at or close to the point of application (POA) when the concentration of monoclonal Ig is high (≥ 2 g/dL). This is more often observed with a polymerized IgM pentamer or an IgA dimer or trimer, as these can further aggregate, than with a monomeric IgG. At a high concentration and with reduced solubility due to polymerization, the monoclonal Ig that did not react with the nonspecific antiserum often does not wash completely out of the gel and is stained. This results in a false band appearing in each track that is not

specific for the monoclonal Ig. Interesting to note is that this false band may be missing in the G track because the sample is twice as dilute as in the remaining tracks. False bands can be easily identified as they are slightly wider than the antisera track itself and therefore wider than true immunofixed bands.

IgM can also trap other Ig's. The trapped Ig forms a complex with its antiserum, but appears at the migration position of IgM. Cryoglobulins, due to their poor solubility, may also cause false bands. When interpretation is ambiguous, repeat the IF after appropriately treating the sample: dilute high levels of Ig, depolymerize polymers or solubilize cryoglobulins.

- At very high concentrations of monoclonal Ig (>3 g/dL) prozoning may occur. This will appear as a band with an unstained hollow in the middle of it similar to an oval doughnut or two close bands.

- Bence Jones Protein (BJP) may bind to other proteins, particularly those with a terminal SH group, e.g., transthyretin, albumin, α -1 antitrypsin, transferrin and enzymes such as lactate dehydrogenase. The band(s) of these complex(es) appear in addition to the major BJP band itself. Treatment with 2ME dissociates the complex. Complexes of α -1 antitrypsin are often seen in patients on chemotherapy.

- Uncommonly, IgM/ κ or IgM/ λ may complex with polyclonal λ or κ , respectively. Then, one band is present in the M, K and L tracks. These bands are at the same migration distance and the non-monoclonal light chain band is weaker. The same phenomenon has been also observed with monoclonal IgG, but so far, not with IgA. When further analysis is required, the use of Fluidil will dissociate the complex in most cases. When IgM is involved, the sample

should be treated with a mixture of Fluidil/2ME before repeating the IF.

- It has been observed in a small number of cases that plasma fibrinogen could complex with polyclonal IgG or polyclonal free light chains. As a result, a faint band can be seen in G, K and L tracks or, in some cases, only in the light chain tracks at the same migration position as the heavy band of fibrinogen in the reference track. Serum contaminated with plasma is unlikely to show this phenomenon.

Other atypical cases

Monoclonal immunoglobulin generally migrates in the same relative position regardless of the procedure used: HYDRAGEL Protein(e), HYDRAGEL β 1- β 2 and HYDRAGEL 2/4 IF. Rarely, monoclonal IgM has been observed to migrate more anodic on HYDRAGEL 2/4 IF gels than on HYDRAGEL Protein(e) or β 1- β 2 gels. Two factors are involved: (i) slightly different composition in the additives of the individual gels and (ii) the mobility of monoclonal IgM tends to be concentration dependent, i.e., neat serum is used with the Protein(e) and β 1- β 2 gels while diluted serum is used for IF analysis.

Several cases have been observed, all including monoclonal IgM, showing a single band in the M, K and L track each that migrated at the same level. It appeared as if one clone of B-cells was producing not only a monoclonal IgM, but also both kappa and lambda light chains - a biological impossibility. Although these were rare cases, they merit further description to illustrate the occasional complexity of monoclonal gammopathies. Three different underlying mechanisms have been identified:

- (i) A biclonal gammopathy producing IgM/ κ and IgM/ λ that happened to co-migrate (in the observed case the heavy chain band was heavier than either of the light chain bands). High Resolution electrophoresis/immunofixation was able

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There's no such thing as spring, summer or winter break for the core lab of a Wayne State University-affiliated teaching hospital. In fact, the tests keep being piled on.

St. John Hospital and Medical Center, part of Detroit's St. John Health System, had terrific success with marketing efforts to generate additional work for its Rapid Response Laboratory. But there was a down side: this newfound cornucopia put a great deal of strain on an already lean lab staff.

"Our previous (manual) methodology worked fine when we were processing 18 patient samples for serum protein electrophoresis a day," says Edward Bessette, technical specialist for the lab. "But suddenly that number escalated to 35 or 40 — and that's a new ballgame entirely."

Through industry contacts, Bessette and his associates were already familiar with a couple of automated electrophoresis systems; they decided it was time to evaluate their options. Sebia's HYDRASYS® system and a competing solution were put to the test — and the HYDRASYS came out ahead.

Designed as a 'walkaway' system, the HYDRASYS expedites the many tedious steps involved in electrophoresis and immunofixation, automatically handling everything from sample application to



Sebia Electrophoresis Gets High Marks From Teaching Hospital
St. John Hospital and Medical Center, Detroit, MI

migration to incubation to staining, destaining and drying.

Sebia's method of sample application scored big points with lab personnel because it relies on a wick instead of the traditional well. "With the HYDRASYS, the sample stays absorbed to the wick, so we don't run the risk of evaporation — which is a common problem with using wells," Bessette explains. "Now we can load one set of 15 samples, put them in the fridge while we load another batch, and not worry about them drying up in the meantime."

Bessette was also impressed with the gel quality of the HYDRASYS — not only for its clarity, but consistency as well. "Because the sample application does not rely on

human skill, we get excellent uniformity of patterns from plate to plate, operator to operator," he notes. "Now our people can focus their attention on other tasks instead of

gels. "Band resolution with Sebia far surpasses our previous system," says St. John staff technologist Gloria Witczak.

Besides making lab techs happy, HYDRASYS also enabled St. John to successfully absorb their huge increase in workload without missing a beat. Now, instead of covering six patients in one run, the lab can test 28 patients per batch. "We can comfortably complete all the day's work without rolling it over to the next shift," Bessette says. "This enhances our service and reliability in the minds of physicians and other customers."

And, as a byproduct of the outstanding clarity and consistency of HYDRASYS results, the lab has decreased the number of repeats — and thereby lowered its material costs. "The instrument allows us to get it right the first time, so we're generating less waste and saving money on gels," Bessette notes.

But in his view, the most important savings a lab can reap from this instrument is the elimination of wasted time. "Tech time is the most valuable commodity we have today...and it's also the most scarce," he says. "Now you have an opportunity to automate a laborious process, conserve manpower and at the same time reduce the chance of human error. Take it."

"Hands-off' gel processing means technologists don't have direct contact with solvents that pose a health hazard."

hazard."

—Edward Bessette, technical specialist, Rapid Response Laboratory

processing plates. We're making better use of their time." (And enhancing their safety too, he adds: 'hands-off' gel processing means technologists don't have direct contact with methanol or other solvents that pose a health hazard.) In terms of gel resolution, Sebia clearly exceeded the lab's expectations; having been accustomed to edge effects and poor quality separation between hemoglobin A and hemoglobin F with previous methods, technologists were pleasantly surprised at the crispness of the separation on Sebia's alkaline and acid



"...technologists were pleasantly surprised at the crispness of the separation on Sebia's alkaline and acid (hemoglobin) gels."

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Thank You for Your Assistance

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